Chordoma Foundation Cell Line Validation U-CH19

Cell morphology, growth and Brachyury expression analysis
February 20, 2020



Validation Report

Cell line: U-CH19

Growth conditions

Media: 4:1 IMDM: RPMI1640 +10% FCS, L-glutamine, Pen/Strep

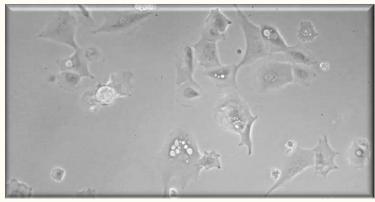
- Change medium every 4-5 days
- Do not passage cells before reaching 90-100% confluency!
- Max. split ratio 1:2 (growing speed is cell density dependent)
- Cell expansion: 2x 25cm² flasks (95% confluent) → 1x 75cm² flask

2x 75cm² flasks (95% confluent) → 1x 175 or 225cm² flask

Percentage viable cells after thawing: >95%

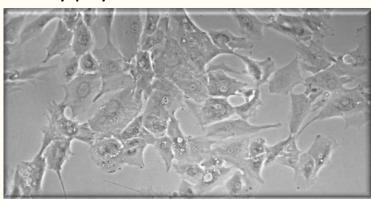
Morphology¹:

mainly physaliferous



Cells 24 hours post thawing (>90% adherent)

mainly physaliferous

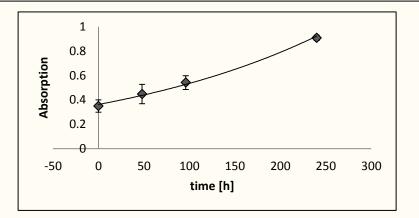


Cells 7 days post thawing

Population doubling time²:

approx. 7 days

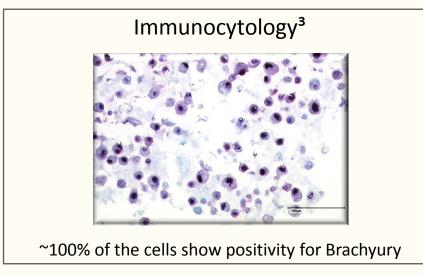
(at recommended density of 5000-7500 cells per cm²)

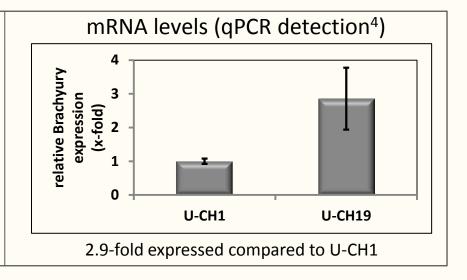


STR profile:

	AMEL	D13S317	D7S820	D16S539	Penta E	THO1	D18S51	D3S1358	D8S1179	TPOX	CSF1PO	Penta D
U-CH19	ХҮ	12	9 10	11 13	12	9 9.3	13 14	14	10 15	8 12	10 12	13

Brachyury expression:





<u>Validation result</u>: The cell line meets the criteria for being a chordoma cell line.

- 1. Cell morphology was monitored and documented using an invert phase contrast microscope. Typical physaliferous morphology of the cells may vary between different cell lines.
- 2. Population doubling was estimated by seeding the cells in recommended densities in 96 well plates. Measurement of viable cells was performed using MTS assays at several time points (n=6 per time point).
- 3. Nuclear positivity for Brachyury was tested using standard immunocytochemistry sections of FFPE cell bocks were used. Stainings were performed using a rabbit monoclonal Brachyury antibody, anti rabbit antibodies and either red or pink or brown dyes.
- 4. Relative T gene expression was calculated using the $\Delta\Delta$ CT method (fold change in relation to T expression of U-CH1).